

Procalcitonin, a new diagnostic and prognostic marker for severe infections

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INTRODUCTION

Morbidity and mortality attributed to severe bacterial infections are still major complications in modern medicine. Early recognition and prompt treatment can reduce morbidity and mortality. Better comprehension of the pathogenesis of sepsis and the response of the host organism have led to new definitions of sepsis (Table 1) [1]. Recent studies suggest that the systemic inflammatory response syndrome (SIRS), sepsis, severe sepsis and septic shock represent different stages of the inflammatory response to infection [2].

Clinical diagnosis based on heart rate, respiratory rate, hypotension, fever, prostration and mental confusion is often difficult and non-specific. Parameters such as white blood cell (WBC) count, shift to the left in differential count, elevated C reactive protein (CRP) or increased erythrocyte sedimentation rate (ESR) can be useful, but are often not specific or sensitive enough. Table 2 shows the sensitivity, specificity and positive and negative predictive values for these tests. Cultures are often negative due to prior antimicrobial treatment or because adequate material is not readily available. The low sensitivity of blood cultures in the diagnosis of severe infections (range 17–69%) has led to the definition of culture-negative sepsis. This broad range in sensitivity is due to differences in defining and

classifying the severity of infection. In a large prospective study, the outcome in patients with culture-negative septic shock was similar to that in patients with microbiologically proven septic shock [2]. Van Griethuysen et al reported a sensitivity and specificity of 53% and 83%, respectively, for temperature >38.5°C [3]. In our study population of 337 episodes, we calculated a sensitivity and specificity of 40% and 65%, respectively, using the threshold suggested by van Griethuysen et al [3] (unpublished data). The positive and negative predictive values for fever are given in Table 2.

The position of cytokines in the diagnosis and prognosis of severe infections is not yet defined and is controversial. Interleukin-6 (IL-6) is found earlier than CRP in severe infections, but it is not sensitive enough to serve alone in the diagnosis of severe infections [4,5]. Tumor necrosis factor (TNF) is known to rise early during acute bacterial infection, but declines rapidly after a peak of only a few hours, resulting in a low sensitivity [4,6]. There are few data available on other diagnostic parameters in the diagnosis of sepsis, such as phospholipase A₂, interferon- γ , neopterin or soluble CD14; too few episodes have been studied to allow definite conclusions, and often complicated laboratory procedures are required [6–9].

PROCALCITONIN

The CALC-I gene encoding the polypeptide hormone calcitonin was one of the first examples of tissue-specific expression of mRNA transcripts [10]. The human CALC-I gene contains six exons [11]. Its predominant product calcitonin, which regulates Ca²⁺ metabolism, is produced by thyroid C-cells, whereas other related peptides are found in neuronal cells. Calcitonin is generated by proteolytic separation

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Table 1 Definitions of the Consensus Conference of the American College of Chest Physicians [1]**Systemic inflammatory response syndrome (SIRS)**

Two or more of the following:

1. Temperature $>38^{\circ}\text{C}$ or $<36^{\circ}\text{C}$
2. Heart rate >90 beats/min
3. Respiratory rate >20 breaths/min
4. White blood cell count $>12.0 \times 10^9/\text{L}$, $<4.0 \times 10^9/\text{L}$, or >0.1 immature forms (bands)

Sepsis

SIRS plus a documented infection (positive culture for organism)

Severe sepsis

Sepsis associated with organ dysfunction, hypoperfusion abnormalities, or hypotension. (Hypoperfusion abnormalities include, but are not limited to, lactic acidosis, oliguria, or an acute alteration of the mental status)

Septic shock

Sepsis-induced hypotension despite fluid resuscitation plus hypoperfusion abnormalities

Culture-negative sepsis

SIRS plus empirical antibiotic treatment for a clinically suspected infection in which all cultures were negative

Culture-negative severe sepsis

SIRS associated with organ dysfunction, hypoperfusion abnormalities, or hypotension

Culture-negative septic shock

SIRS associated with hypotension despite fluid resuscitation plus hypoperfusion abnormalities. All cultures were negative, yet empirical antibiotic treatment for a clinically suspected infection was prescribed

Table 2 Sensitivity and specificity and negative and positive predictive value of temperature, WBC count, ESR, CRP, blood cultures and procalcitonin (PCT) in patients with sepsis

Text	Reference	Sensitivity (%)	Specificity (%)	Positive predictive value (%)	Negative predictive value (%)
Blood culture	[2]	17–69	NA	NA	NA
	[42]	63	NA	NA	NA
Temperature $>38.5^{\circ}\text{C}$	[3]	53	83	43	88
	UD	40	65	59	46
WBC $>12 \times 10^9/\text{L}$	[3]	73	32	18	85
	UD	55	67	29	86
ESR >30 mm/h	[43]	70	55	43	78
CRP >10 mg/L	[43]	67	69	51	81
	UD	42	68	38	72
PCT >0.5 ng/mL	[24]	0	79	61	78

UD, unpublished data; NA, not available.

from the larger prehormone, procalcitonin (PCT), which consists of 116 amino acids [12]. These peptides have already been used as markers in medullary thyroid carcinoma and in a number of other malignant processes [13–16]. In some of these cases, particularly lung cancer, PCT is found to be elevated without the presence of mature calcitonin. However, the biological role of this mechanism and the function of PCT remain unknown. The first hints about a mitogenic activity of PCT on human osteoblastic cells led to speculations about its use in patients with osteoporosis [17–20].

In healthy volunteers, PCT serum concentrations are below 0.15 ng/mL, although there may be a peak in the first day of life which is independent of an infectious stimulus [21,22]. The threshold value of

PCT in patients with SIRS and suspected sepsis seems to be 0.5 ng/mL [23,24].

PCT is determined semi-automatically with an immunoluminometric assay using antibody-coated tubes in a luminometer. The test is specific for PCT, with a detection limit of 0.1 ng/mL and good reproducibility. The assay uses two monoclonal antibodies. One is a capture antibody directed against the 96–106 sequence in PCT, and the other is a tracer antibody directed against the 70–76 residues [21]. At room temperature a reduction in PCT plasma concentrations of 12.3% per 24 h is reported. There seems to be no significant influence of the blood sampling technique (arterial or venous line) or of repeated freezing–thawing cycles on the PCT concentrations [25].

As calcitonin and calcitonin-related peptides have been found in human neuroendocrine lung cells [26,27], efforts were first made to determine PCT in patients with inhalatory injuries following burns [28,29]. Here it is found at high concentrations without the presence of mature calcitonin. Higher concentrations of PCT correlated with mortality but not always with the burnt body-surface area. In some instances PCT remains elevated even after spirometry measurements return to normal values, indicating a long-term response of pulmonary neuroendocrine cells to burns [30]. In the same study, PCT and IL-6 concentrations were not associated with smoke inhalation or infection.

To investigate the role of PCT in severe infections, Dandona et al measured PCT concentrations in healthy volunteers after the injection of endotoxins derived from *Escherichia coli* [21]. TNF- α levels peaked very early after 90-min and fell to baseline levels after 6 h. IL-6 increased more gradually, peaking after 3 h and reaching the baseline concentration after 8 h. PCT peaked after 6 h and remained at a plateau for more than 24 h. In patients with bacterial and aspiration pneumonia, PCT is found to be only moderately elevated, correlating with the radiographic changes [31,32]. Values of up to 2 ng/mL are found on the first days of an episode.

There are only a few clinical studies investigating the position of PCT in the diagnosis and prognosis of infections. Assicot et al [33] were the first to report PCT serum levels in 79 pediatric patients; 19 of them had severe bacterial infection, with serum PCT concentrations ranging from 6 to 53 ng/mL. Patients with local infection or viral infection had lower values. They reported a close relation with infectious complications. We prospectively studied 337 adult patients admitted to an internal medicine department with suspected infection [24]. Patients who were classified as having SIRS had significantly lower PCT levels than patients with SIRS and in addition infection or septicemia or septic shock (Table 3). Other authors have found

PCT levels above 1 ng/mL only in patients with septic shock. The causative organism (whether a Gram-negative or Gram-positive bacterium, or another micro-organism such as a *Plasmodium* species or fungus) does not significantly influence the level of PCT in serum. Also, episodes with culture-negative sepsis are characterized by elevated PCT concentrations [34]. Immunocompromised patients (HIV infection or malignant diseases) also appear to show high serum PCT concentrations during sepsis, but leukopenia seems to be associated with lower PCT values after day 2 of the sepsis episode [23,24]. It has been reported that PCT may serve as a useful marker for the detection of systemic bacterial infection in patients with systemic autoimmune disease [35]. In the same study no correlation was seen between the degree of renal impairment and PCT concentrations.

Data are also available on PCT concentrations in neonatal infections. There seems to be a normal peak PCT concentration in healthy newborns in the first few hours after birth, soon declining to baseline levels. The available data suggest that in neonates with severe infections PCT levels rise more rapidly than CRP levels and that the sensitivity is higher [22]. In pediatric patients PCT seems to distinguish better between bacterial and viral meningitis than CRP or an assay of cells and protein in cerebrospinal fluid [33,36].

There is limited information on PCT concentrations after surgical procedures. Efforts have been made to use PCT in the diagnosis of acute rejection of heart transplants. Patients with acute cellular rejection of the transplanted heart showed no circulating PCT; however, patients with bacterial or fungal infection showed moderate to high levels of PCT [37].

Elevated PCT levels have also been reported in tropical diseases. In patients with melioidosis due to *Burkholderia pseudomallei*, initial high PCT values have been reported to have prognostic value for higher mortality [38]. In Asian as well as European populations with malaria, PCT is elevated, reaching values over 100 ng/mL. This seems to correlate with severity. In the first few hours of infection, PCT values reach peak levels, declining rapidly under therapy [39,40].

Few data are available on the prognostic usefulness of PCT in patients with severe infections. In a study on 30 intensive care unit patients with sepsis or septic shock, PCT seemed to be higher in non-surviving patients than in survivors. CRP, TNF- α , neopterin, and IL-6 were not significantly correlated with the outcome of the infection [41]. In our study of 337 episodes, PCT levels were followed for 9 days. On admission, the mean PCT value for survivors was 4.4 ng/mL, whereas for non-survivors the mean values was 15.2 ng/mL ($p=0.002$).

Table 3 Maximum PCT values from days 0 and 1 in 337 patients with suspected sepsis [24]

Group	Mean PCT (ng/mL)	Standard deviation	No. of cases
1. SIRS only	0.6	2.2	215
2. SIRS + infection, microbiologically proven	6.6	22.5	53
3. SIRS + septicemia	8.5	19.0	49
4. SIRS + septic shock	34.7	68.4	20
Entire population	4.7	21.6	337

CONCLUSION

Although many tests are available in the diagnosis of severe infection, there is a need for more sensitive and specific diagnostic tools. The monitoring of success or failure of anti-infective therapy, especially in critically ill patients, is still unsatisfactory. Many studies have independently shown that PCT is elevated early in episodes of severe infections. However, there are large interindividual differences which seem not to be related to the disease. Differences in the definition of sepsis and severe infection may lead to conflicting results.

There are few statistically significant data on the threshold of PCT in diagnosis of severe infections, as most studies do not contain enough cases to lead to statistically established statements. The normal values for PCT in a healthy population are well established, but there are not enough studies on PCT values in patients with non-infectious diseases. The prognostic value of PCT needs to be established in larger study populations. The number of cases studied so far is limited, and it seems that if PCT is initially elevated and does not decline in the course of infection the mortality is higher. This is also the case in patients who do not respond to antimicrobial treatment. Multicenter studies are required to resolve these issues. The mechanism of elevated serum PCT levels in patients with severe infections is not well understood. However, it may be useful to elucidate the role of PCT in the pathologic mechanisms in severe infections, as this may lead to better utilization of this promising laboratory parameter.

References

1. American College of Chest Physicians–Society of Critical Care Medicine Consensus Conference. Definitions for sepsis and organ failure and guidelines for the use of innovative therapies in sepsis. *Crit Care Med* 1992; 20: 864–75.
2. Rangel-Frausto S, Pittet D, Costigan M, Hwang T, Davis CS, Wenzel RP. The natural history of the systemic inflammatory response syndrome (SIRS). *JAMA* 1995; 273: 117–23.
3. van Griethuysen D, Bergmans D, Bonten M, et al. The value of white blood cell count and differential count in patients admitted to the ICU with and without infections [abstract J98]. In: Program and abstracts of the 35th International Conference on Antimicrobial Agents and Chemotherapy, San Francisco, Washington DC: American Society for Microbiology, 1995: 274.
4. Damas P, Ledoux D, Nys M, et al. Cytokine serum level during severe sepsis in human IL-6 as a marker of severity. *Ann Surg* 1992; 215: 356–62.
5. Rintala E, Pulkki K, Mertsola J, Nevalainen T, Nikoskelainen J. Endotoxin, interleukin-6 and phospholipase-A2 as markers of sepsis in patients with hematological malignancies. *Scand J Infect Dis* 1995; 27: 39–43.
6. Blanco A, Solis G, Arranz E, Coto GD, Ramos A, Telleria J. Serum levels of CD14 in neonatal sepsis by Gram-positive and Gram-negative bacteria. *Acta Paediatr* 1996; 85: 728–32.
7. Jurgens ES, Henderson DC. Inflammatory and immunological markers in preterm infants: correlation with disease. *Clin Exp Immunol* 1996; 105: 551–5.
8. Nyman KM, Uhl W, Forsstrom J, Buchler M, Beger HG, Nevalainen TJ. Serum phospholipase A2 in patients with multiple organ failure. *J Surg Res* 1996; 60: 7–14.
9. Rintala EM, Nevalainen TJ. Group II phospholipase A2 in sera of febrile patients with microbiologically or clinically documented infections. *Clin Infect Dis* 1993; 17: 864–70.
10. Rosenfeld MG, Mermod JJ, Amara SG, et al. Production of a novel neuropeptide encoded by the calcitonin gene via tissue-specific RNA processing. *Nature* 1983; 304: 1129–35.
11. Amara SG, Evans RM, Rosenfeld MG. Calcitonin/calcitonin gene-related peptide transcription unit: tissue-specific expression involves selective use of alternative polyadenylation sites. *Mol Cell Biol* 1984; 4: 2151–60.
12. LeMoullec JM, Jullienne A, Chénais J, et al. The complete sequence of human preprocalcitonin. *FEBS Lett* 1984; 167: 93–7.
13. Born W, Beglinger C, Fischer JA. Diagnostic relevance of the amino-terminal cleavage peptide of procalcitonin (PAS-57), calcitonin and calcitonin gene-related peptide in medullary thyroid carcinoma patients. *Regul Pept* 1991; 32: 311–9.
14. Ghillans P, Motte P, Troden F. Identification and measurement of calcitonin and precursor molecules of patients with malignant diseases. *Cancer Res* 1989; 49: 6845–51.
15. Raue F, Blind E, Grauer A. PDN-21 (katecalcine) and chromogranin A: tumor markers for medullary thyroid carcinoma. *Henry Ford Hosp Med J* 1992; 40: 296–8.
16. Schlumberger M, Gicquel C, Lumbroso J, et al. Malignant pheochromocytoma: clinical, biological, histologic and therapeutic data in a series of 20 patients with distant metastases. *J Endocrinol Invest* 1992; 15: 631–42.
17. Burns DM. Procalcitonin, a bone cell mitogene. *Proc Natl Acad Sci USA* 1989; 86: 9519–23.
18. Cotton P. Peptide portions may hold the key to amplifying bone against porosis. *JAMA* 1990; 263: 621.
19. Guenther HL, Fleisch H. The procalcitonin amino-terminal cleavage peptide (N-proCT) lacks biological activity on normal clonal rat osteoblastic and preosteoblastic cells in vitro. *Calcif Tissue Int* 1991; 49: 138–40.
20. Hassager C, Bonde SK, Anderson MA, Rink H, Spelsberg TC, Riggs BL. Procalcitonin NH₂-terminal cleavage peptide has no mitogenic effect on normal human osteoblast-like cells. *J Bone Miner Res* 1991; 6: 489–93.
21. Dandona P, Nix D, Wilson MF, et al. Procalcitonin increase after endotoxin injection in normal subjects. *J Clin Endocrinol Metabol* 1994; 79: 1605–8.
22. Monneret G, Labaune JM, Isaac C, Bienvenu F, Putet G, Bienvenu J. Procalcitonin and C-reactive protein levels in neonatal infections. *Acta Paediatr* 1997; 86: 209–12.
23. Gerard Y, Hober D, Assicot M, et al. Procalcitonin as a marker of bacterial sepsis in patients infected with HIV-1. *J Infect* 1997; 35: 41–6.

24. Al-Nawas B, Krammer I, Shah PM. Procalcitonin in the diagnosis of severe infections. *Eur J Med Res* 1996; 1: 331-3.
25. Meisner M, Tschakowsky K, Schnabel S, Schmidt J, Katalinic A, Schüttler J. Procalcitonin—influence of temperature, storage, anticoagulation and arterial or venous asservation of blood samples on procalcitonin concentrations. *Eur J Clin Chem Clin Biochem* 1997; 35: 597-601.
26. Becker KL, Snider RH, Silva OL, Moore MF. Calcitonin heterogeneity in lung cancer and medullary thyroid cancer. *Acta Endocrinol* 1978; 89: 89-99.
27. Becker KL, Monaghan KG, Silva OL. Immunocytochemical localisation of calcitonin in Kulschatzky cells of human lung. *Path Lab Med* 1980; 104c: 196-8.
28. Nylen ES, O'Neill W, Jordan MH, et al. Serum procalcitonin as an index of inhalation injury in burns. *Horm Metab Res* 1992; 24: 439-42.
29. Nylen ES, Jeng J, Jordan MH, et al. Late pulmonary sequela following burns: persistence of hyperprocalcitonemia using a 1-57 amino acid N-terminal flanking peptide assay. *Respir Med* 1995; 89: 41-6.
30. Carsin H, Assicot M, Feger F, et al: Evolution and significance of circulating procalcitonin levels compared with IL-6, TNF α and endotoxin levels early after thermal injury. *Burns* 1997; 23: 218-24.
31. de Werra I, Jaccard C, Corradin SB, et al. Cytokines, nitrite/nitrate, soluble tumor necrosis factor receptors, and procalcitonin concentrations: comparisons in patients with septic shock, cardiogenic shock, and bacterial pneumonia. *Crit Care Med* 1997; 25: 607-13.
32. Nylen ES, Snider RH Jr, Thompson KA, Rohatgi P, Becker KL. Pneumonitis-associated hyperprocalcitoninemia. *Am J Med Sci* 1996; 312: 12-8.
33. Assicot M, Gendrel D, Carsin H, Raymond J, Guilbaud J, Bohuon C. High serum procalcitonin concentrations in patients with sepsis and infection. *Lancet* 1993; 341: 515-8.
34. Al-Nawas B, Shah PM. Procalcitonin in patients with and without immunosuppression and sepsis. *Infection* 1996; 24: 434-6.
35. Eberhard OK, Haubitz M, Brunkhorst FM, Kliem V, Koch KM, Brunkhorst R. Procalcitonin for differentiation between activity of systemic autoimmune disease and invasive bacterial infection. *Arthritis Rheum* 1997; 40: 1250-6.
36. Gendrel D, Raymond J, Assicot M, et al. Measurement of procalcitonin levels in children with bacterial or viral meningitis. *Clin Infect Dis* 1997; 24: 1240-2.
37. Staehler M, Hammer C, Meiser B, Reichart B. Procalcitonin: a new marker for differential diagnosis of acute rejection and bacterial infection in heart transplantation. *Transplant Proc* 1997; 29: 584-5.
38. Smith MD, Suputtamongkol Y, Chaowagul W, et al. Elevated serum procalcitonin levels in patients with melioidosis. *Clin Infect Dis* 1995; 20: 641-5.
39. Al-Nawas B, Shah PM. Procalcitonin in acute malaria. *Eur J Med Res* 1997; 2: 206-8.
40. Davis TM, Assicot M, Bohuon C, St John A, Li GQ, Anh TK. Serum procalcitonin concentrations in acute malaria. *Trans R Soc Trop Med Hyg* 1994; 88: 670-1.
41. Oberhoffer M, Bögel D, Morgner C, Vogelsang H, Koenigs D, Reinhart K. Procalcitonin is higher in non-survivors at day one of the diagnosis of sepsis and severe sepsis/septic shock. 1. International Jena Symposium. *Clin Intens Care* 1996; 7: 46.
42. Shah PM, Schaumann RF. Positivitätsrate von Blutkulturen bei Sepsis. *J Lab Med* 1997; 21: 120-1.
43. Katz PR, Gutman SI, Richman G, Karuza J, Bartholomew WR, Baum J. Erythrocyte sedimentation rate and c-reactive protein compared in the elderly. *Clin Chem* 1989; 35: 466-8.